In the specification:

Please cancel previous Figure 7D.

Please replace the description of Figure 2 in the Brief Description of the Drawings (as amended on June 19, 2001) with the following:

-Figure 2: Nucleotide sequence of the cDNA of IL-13Rβ. (SEQ ID NO. 1), and comparison of the protein sequences of IL-5R (SEQ ID NO. 5) and IL-13Rβ (SEQ ID NO. 2).

FIG. 2A & 2B   nucleotide sequence of the cDNA of IL-13Rß (SEQ ID NO. 1). The amino acids corresponding to the deduced signal peptide of the nucleic sequence are indicated in italics and those corresponding to the transmembrane domain are indicated in bold characters. The potential N-glycosylation sites (Asn-X-Ser/Thr) are underlined;

FIG. 2C & 2D   alignment of the amino acids of the IL-13Rß (SEQ ID NO. 2) and IL-5R (SEQ ID NO. 5) sequences. The protein sequences of IL-13R (SEQ ID NO. 2) and IL-5R (SEQ ID NO. 5) are aligned as described above (24). The cysteine residues and the WSXWS (SEQ ID NO. 13) motif which are characteristic of this family of receptors are boxed.

Please replace the description of Figure 7 in the Brief Description of the Drawings (as amended on June 19, 2001) with the following:

-Figure 7: nucleotide sequence of the IL-13Rβ (SEQ. ID NO. 3) cDNA and comparison of the protein sequences of human IL-13Rβ (SEQ. ID NO. 4) and of murine IL-13Rβ (SEQ. ID NO. 6).

FIG. 7A, 7B, 7C & 7D &~7B Nucleotide sequence of the IL-13Rβ (SEQ. ID NO. 3) cDNA. The amino acids corresponding to the signal peptide deduced from the nucleic sequence are underlined with a dotted line and those corresponding to the transmembrane domain are underlined with a double line. The potential N-glycosylation sites (Asn-X-Ser/Thr) are boxed.

FIG. 7E & 7F 7C~&~7D Alignment of the amino acids of human IL-13Rβ (SEQ. ID NO. 4) and of murine IL-13Rβ (SEQ. ID NO. 6). The protein sequences of human IL-13Rβ (SEQ. ID NO. 4) and of murine IL-13Rβ (SEQ. ID NO. 6) are aligned as described above (24). The cysteine residues and the motif WSXWS (SEQ. ID NO. 3) which are character-istic of this family of receptors are boxed.

Please replace the paragraph beginning on page 23, line 21, and continuing onto page 24, (as amended on October 21, 1999) with the following replacement paragraph:

The strategy for the cloning and expression which was used has been previously described (17). A cDNA library containing 2×10^5 recombinant clones was constructed (26) using Caki-1 cells. The library was divided into batches of 1000 cDNAs in which the DNA of each batch, in plasmid form, was introduced into COS-7 cells (29). The binding of labelled IL-13 to the transfected COS-7 cells makes it possible to identify the batches of clones encoding an IL-13 receptor. The positive batches were distributed out and rescreened until a single clone capable of carrying out the synthesis of a cell surface protein capable of binding IL-13 is identified. Two independent IL-13Rβ cDNAs were finally isolated. The complete nucleotide

sequence of the IL-13Rβ cDNA and the amino acid sequence
deduced therefrom are shown in Figures 2a and 2b. The
cDNA has a length of 1298 bases excluding the poly-A tail
and has a short 3' untranslated region of 106 bases. A
canonical AATAAA (SEQ ID NO. 14) polyadenylation signal
is in the expected place. The open reading frame between
nucleotides 53 and 1192 defines a polypeptide of 380
amino acids. The sequence encodes a membrane protein with
a potential signal peptide, a single transmembrane domain
and a short intracytoplasmic tail.

Please replace the paragraph beginning on page 24, line 17 with the following
replacement paragraph:

Alignment studies demonstrate homologies with the
human IL-5R α chain (51% similarity and 27% identity,
Figures 2c and 2dFigure 2b) and, to a lesser extent, with
the prolactin receptor. It is interesting to note that
the IL-5R complex consists of an α chain which binds IL-5
but which needs another protein, the β chain shared with
the IL-3 and GM-CSF receptors, to form a high-affinity
receptor which is capable of transducing a signal (31).

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